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Deeper knowledge of shallow waters: reviewing the invertebrate fauna of southern African temporary wetlands

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- 1 Deeper knowledge of shallow waters: reviewing the invertebrate fauna of
- 2 southern African temporary wetlands
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Abstract

| 21 | Temporary lentic wetlands are becoming increasingly recognised for their collective role in |
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| 22 | contributing to biodiversity at the landscape scale. In southern Africa, a region with a high |
| 23 | density of such wetlands, information characterising the fauna of these systems is disparate |
| 24 | and often obscurely published. Here we provide a collation and synthesis of published |
| 25 | research on the aquatic invertebrate fauna inhabiting temporary lentic wetlands of the region. |
| 26 | We expose the poor taxonomic knowledge of most groups, which makes it difficult to |
| 27 | comment on patterns of richness and endemism. Only a few groups (e.g. large branchiopods, |
| 28 | ostracods, copepods and cladocerans) appear to reach higher richness and/or endemicity in |
| 29 | temporary wetlands compared to their permanent wetland counterparts. IUCN Red List |
| 30 | information is lacking for most taxa, thus making it difficult to comment on the conservation |
| 31 | status of much of the invertebrate fauna. However, except for a few specialist groups, many |
| 32 | of the taxa inhabiting these environments appear to be habitat generalists that |
| 33 | opportunistically exploit these waterbodies and this is hypothesized as one of the reasons why |
| 34 | endemism appears to be low for most taxa. Given that taxonomy underpins ecology, the |
| 35 | urgent need for more foundational taxonomic work on these systems becomes glaringly |
| 36 | apparent. |
| 37 | Keywords |
| 38 | aquatic invertebrates; wetland invertebrates; ephemeral wetlands; temporary ponds; |
| 39 | ephemeral ponds; pans; southern Africa; African wetlands |
| 40 | |
| 41 | Introduction |
| 12 | Invertebrates dominate the faunas of temporary wetlands worldwide, in terms of species |
| 43 | diversity, abundance and animal biomass (Williams, 2006). Insects and crustaceans, in |
| 14 | particular, are ubiquitous in such systems. Temporary wetlands house a suite of uniquely |
| 45 | specialized invertebrates (Wiggins et al., 1980; Collinson et al., 1995; Brucet et al., 2005; |
| 46 | Escalera-Vázquez & Zambrano, 2010; Strachan et al., 2015) which are particularly |
| 1 7 | vulnerable to habitat and hydrological modifications (Calhoun et al., 2017; Dalu et al., |
| 18 | 2017a). The southern African region, with its distinct climatic and geomorphic features, has |
| 19 | one of the highest densities of temporary depression wetlands (known locally as 'pans') |
| 50 | worldwide (Goudie & Wells, 1995). Reviews of the invertebrate fauna of temporary wetlands |

51 have been conducted for a number of regions globally (e.g. Batzer et al., 1999; Batzer &

Boix, 2016), but not for southern Africa; as such, this study aims to fill the knowledge gap.

53 Southern Africa is generally defined as the area south of the Kunene River in the west and the

54 Zambezi River in the east (including the countries of Botswana, Lesotho, Namibia, South

Africa, Swaziland, Zimbabwe, and southern parts of Angola and Mozambique). These

boundaries are referred to as 'the region' or 'the southern African region' throughout this

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Providing a checklist of all the species within each invertebrate group known from temporary wetlands in the region is beyond the scope of this review and in many cases insufficient information is available. For certain groups, we summarise reported checklists and highlight gaps in taxonomic and ecological understanding. We review taxonomic and biodiversity studies pertaining to freshwater invertebrates in temporary lentic wetlands in the region, group-by-group. Our primary aim is to provide an overview of available information (taxonomic, faunistic and ecological) for each of the major invertebrate taxa found in temporary lentic wetlands in the region, highlighting major gaps in knowledge. The key adaptations that each invertebrate taxon has evolved to cope with desiccation are also briefly covered where such information exists. For general information on wetland invertebrate adaptations to drought, see Wiggins et al. (1980), Brendonck & De Meester (2003), Williams (2006), Strachan et al. (2015) and Batzer & Boix (2016).

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Methods

- A comprehensive review of all published material in peer-reviewed journals for the period
- 73 1960 to 2016 was conducted using a combination of the following terms: 'temporary',
- 74 'seasonal', 'ephemeral', 'vernal', 'endorheic', 'rock', 'rain', 'pond', 'pool', 'pan', 'tarn',
- 75 'depression', 'southern Africa', plus all the individual countries within it, and 'invertebrates',
- 76 plus all the major taxa within. References to other important studies cited in the published
- 77 material above were also referred to and in turn these were also searched thoroughly for
- further important cited work. In this way, the relevant literature has been comprehensively
- surveyed. Only those papers that tackled invertebrate populations or communities from
- wetland systems that temporarily dry up were included in the final list used for this review.
- However, in certain cases, there was no mention of whether the wetland was temporarily
- 82 inundated or not. In such cases it was necessary to pay careful attention to the mention of or

inference to flagship taxa (sensu De Roeck et al., 2007) or the adaptive traits to drying conditions (Lahr, 1997) as this would confirm the temporary nature of the wetland. The review aims to focus on lentic isolated depressional wetlands that are not connected to longitudinal watercourses (i.e. non-riverine systems). However, such specific habitat information is often not available for a given taxon, and thus we include taxa that have simply been recorded as occurring in temporary wetland environments, even if information on the specific nature of these environments is not available for the taxon in question.

The material from peer-reviewed journals was further complemented by consulting key texts in the 'grey literature' that contain comprehensive species lists, including the Water Research Commission's 'Guides to the Freshwater Invertebrates of Southern Africa' (relevant chapters are referenced separately throughout the text). For the molluscs, for example, Brown's (1994) comprehensive account of African freshwater gastropods was used to systematically search each species' information to establish whether or not it is known to inhabit temporary wetland environments. Similarly, for odonates, which have several species checklist publications covering the various sub-regions of southern Africa (Martens et al., 2003; Kipping, 2010; Samways & Simaika, 2016), it was assessed whether the habitat preferences for the various species have been noted or not, and this information was used to construct a list of temporary wetland-dependent species.

Major invertebrate groups

Porifera

Sponges occur in permanent and temporary freshwater systems worldwide, both lotic and lentic (Manconi & Pronzato, 2008). In southern Africa there are two known families, Spongillidae (five genera, seven species) and Potamolepidae (two genera, species information unavailable, Heeg, 2002a). There appears to be a low richness of sponge taxa in southern African freshwater environments compared to other regions (Manconi & Pronzato, 2008) but, as with many of the lesser-known phyla, additional collecting effort is likely to reveal more species (Heeg, 2002a). *Ephydatia fluviatilis* (Linneaus, 1758) of the Spongillidae is the only known species that has often been recorded from temporary wetlands in the region, usually found on the stems of emergent macrophytes. This species appears to be distributed throughout South Africa (SA) and Namibia (Heeg, 2002a). Sponges generally survive

114 drought periods through formation of gemmules, which are internal buds composed of 115 amoebocytes surrounded by an external protective coat comprising collagen layers embedded 116 with spicules (for review, see Frost, 1991). 117 Cnidaria 118 Hydra (class Hydrozoa) is the best-known cnidarian taxon in freshwater wetlands, and is 119 particularly abundant in temporary lentic ponds, where it is represented by solitary polyps 120 (Williams, 2006). In the region, out of the five species of Hydrozoa recorded, only 121 Limnocnida tanganjicae (Günther, 1893) is recognised as indigenous to Africa, the rest being 122 cosmopolitan species (Rayner et al., 2002). An invasive freshwater jellyfish, Craspedacusta 123 sowerbyi (Lankester, 1880), has been recorded in large numbers of man-made reservoirs 124 (Rayner et al., 2002). Hydras are capable of both asexual and sexual reproduction and are 125 known to survive in desiccated wetlands via the latter technique, whereby fertilized eggs are 126 surrounded by a thecal coat that is resistant to external desiccation and extreme changes in 127 environmental conditions in general (Williams, 2006). 128 *Platyhelminthes (flatworms)* 129 'Turbellarians' are common and widespread in southern African temporary wetlands, yet no 130 taxonomic key exists for species or even genera in the region (Appleton, 2002a). Young 131 (1976) recorded 80 valid species of freshwater turbellarians across Africa, but little work has 132 been done since to describe African taxa. Artois et al. (2004) described one new genus and 133 four new species of flatworm from ephemeral rock pools in Botswana. Intensive ecological 134 studies of temporary rock pool invertebrate communities in south-eastern Botswana 135 (Brendonck et al., 2002) found that turbellarians (particularly *Mesostoma* spp.) are key 136 predators of anostracan (fairy shrimp) egg banks in these systems, seriously affecting 137 anostracan population sizes (De Roeck et al., 2005). Mesostoma species have even been 138 recorded preying on adult *Branchipodopsis* shrimp in these pools (Brendonck et al., 2000b). 139 Curtis (1991) reported that only one species of freshwater turbellarian, Mesostoma brincki 140 Marcus 1970, had been positively identified from Namibia. In a comprehensive global 141 collation of turbellarian species with Gondwanan affinities, Noreña et al. (2003) listed six 142 freshwater turbellarians (Xenostenostomum microstomoides Reisinger, 1976; Prorhynchus 143 brincki Marcus, 1955; Bothrioplana semperi Braun, 1881; Mesostoma brincki; M. ewerum 144 Du Bois-Reymond Marcus 1951; *Phaenocora foliacea* (Böhmig, 1914)) from the region, but

145 did not indicate how many of these occur in temporary wetlands. Housen et al. (2014) 146 reported *P. foliacea* from a temporary pond in Hluhluwe Game Reserve, SA. Tubellarians 147 can cope with the periodic drying up of temporary wetlands by producing cysts, and these can 148 come out of dormancy after 5-7 minutes of inundation (Van Steenkiste et al., 2010). 149 The other major platyhelminth groups with notable freshwater representatives are the 150 trematodes and cestodes, both of which are parasitic. There is no existing taxonomic key for 151 African trematodes and they are poorly known. Appleton (2002a) treats them according to 152 groups (types) based on morphology of the cercariae. The obvious exception to an otherwise 153 poorly-known fauna is the genus Schistosoma, which has seven species implicated in human 154 or bovine bilharzia infection and that uses Biomphalaria or Bulinus gastropod snails as 155 intermediate hosts (see 'Molluscs' below for further review of this topic). False tapeworms 156 (subclass Cotyloda) constitute the group of cestodes that can be found in freshwater 157 environments, and in the region are a common parasite of the platanna frog *Xenopus laevis* 158 (Daudin, 1802), but their free-living larval stages are microscopic and are never seen unless 159 specifically sought (Appleton, 2002a). 160 Gastrotricha 161 Gastrotrichs have been recorded from temporary wetlands worldwide, but very little 162 ecological or taxonomic information is available on these organisms in freshwater 163 environments of southern Africa (Williams, 2006). In the review of Heeg (2002b), not a single species description existed for the region. The genera that have been recorded so far in 164 165 the region are cosmopolitan, being found particularly in Europe but also elsewhere (Heeg, 166 2002b). One recent exception is the new planktonic gastrotrich species, Kijanebalola 167 devestiva Todaro, Perissinotto and Bownes, 2013, described from a shallow pond on the 168 shores of iSimangaliso Wetland Park (KwaZulu-Natal, SA). However, despite the pond being 169 small, it appears to be permanently inundated (Todaro et al., 2013). Nevertheless, gastrotrichs 170 are well adapted to life in temporary wetlands through their production of dormant eggs 171 (Ricci & Balsamo, 2000). 172 Bryozoa Bryozoans have been noted as relatively common in freshwater environments in southern 173 174 Africa (Heeg, 2002c; de Moor & Day, 2013), but remain largely uncollected and unstudied in 175 the region. Heeg (2002c) comments that the bryozoan fauna of Africa appears to contain

176 mostly cosmopolitan species, but lists only four species as recorded from the region (Plumatella emarginata (Allman, 1844), P. repens (Linnaeus, 1758), Lophopodella capensis 177 178 (Sollas, 1908) and *Hyalinella punctata* (Hancook, 1850)). It is highly likely that bryozoans 179 inhabit temporary wetlands across large areas of the southern African region, given that they 180 are well adapted to survive desiccation. Bryozoans germinate by producing desiccation-181 resistant statoblasts that have hooks for clinging to large mobile animals such as waterfowl 182 and are thus able to disperse passively over wide geographical areas (Bilton et al., 2001). 183 Nematoda 184 Nematodes are a highly diverse group worldwide (conservatively estimated at one million 185 species, Abebe et al., 2008) and there are approximately 150 known species of freshwater 186 nematodes in southern Africa, of which most are likely to be capable of surviving dry periods 187 in moist soil (Heyns, 2002). Thirty-four of the 150 species listed by Heyns (2002) have been 188 recorded in semi-aquatic environments and are capable of surviving near-terrestrial 189 conditions, for example in moist soil during the dry phase of a temporary wetland. 190 Nematodes have commonly been recorded elsewhere as able to enter a quiescent phase 191 without water loss in response to drying in temporary ponds (Womersley & Ching, 1989; 192 Wharton, 2004; Abebe et al., 2006) and thus are not expected to be limited by extreme 193 environments in southern African temporary wetlands. 194 Nematomorpha 195 Information on the genera and species of the parasite phylum Nematomorpha (horsehair 196 worms) in southern African water bodies is extremely sparse, but the few records that do 197 exist suggest a wide distribution (Rayner & Appleton, 2002). Corrêa (1951) reported two 198 species of *Prostoma* as widespread in SA. In a review of known African species, Schmidt-199 Rhaesa & Perissinotto (2016) reported a further six species from SA belonging to the genera 200 Chordodes, Paragordius and Beatogordius. They also described a previously unconfirmed 201 species (Chordodes ferox Camerano, 1897) collected from a live praying mantid floating in a 202 river in the Eastern Cape province, SA. Nine species recorded by Corrêa (1951) and Schmidt-203 Rhaesa & Perissinotto (2016) appear to be all that is known taxonomically of the fauna from 204 the southern African region to date. Nematomorph larvae are dependent on an aquatic 205 medium, where they generally penetrate the bodies of arthropods (particularly orthopterans) 206 and spend the vast majority of their lives growing into large adult worms inside the host

body. Thus, nematomorphs possess traits that pre-adapt them to life in temporary wetlands
 (Williams, 2006) and it is likely that they inhabit such environments in southern Africa.

Rotifera

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Microscopic animals belonging to the phylum Rotifera (commonly called 'wheel animals') are found in a wide variety of freshwater habitats from large lakes to damp moss and are well adapted to life in temporary wetlands. Rotifers have two major means of dealing with desiccation. Bdelloid rotifers are able to slowly dry out and then return to activity when they are rehydrated, a process known as cryptobiosis (Brain, 2002). Monogonant rotifers instead survive as desiccation-tolerant resting eggs (Ricci & Balsamo, 2000). Both desiccated bdelloids and monogonant eggs are carried by wind and for this reason freshwater rotifers are mostly cosmopolitan, being able to flourish wherever suitable conditions are encountered (Brain, 2002). According to Dumont (1983), African rotifers are particularly cosmopolitan, although no comprehensive species list exists. Brain (2002) provided a list and an identification key for the 47 genera of freshwater rotifers known from southern Africa (8 digononts and 39 monogononts), but it is not known how many of these taxa are found in temporary wetland environments. Anusa et al. (2012) reported eight rotifer species (belonging to the genera Asplanchna, Brachionus, Conochilus, Epiphanes, Lepadella, Rotaria and Synchaeta) as common inhabitants across a range of hydroperiods in temporary rock pools in northern Zimbabwe. They found that rotifers were more abundant in pools with longer hydroperiods, but were able to withstand the ephemerality of the shallow short-lived pools. Dalu et al. (2016) recorded five rotifer species belonging to four genera (Brachionus, Euchlanis, Filinia, Pompholyx) in an ephemeral pond in the Eastern Cape. However, more species are expected to be found with more studies currently underway. Riato et al. (2014) compared the zooplankton communities of temporary and permanent depression wetlands in the Mpumalanga Highveld, SA, and found that *Platyias quadricornis* (Ehrenberg, 1832) only occurred in the temporary wetlands. All nine species reported from the studies of Anusa et al. (2012) and Riato et al. (2014) are widespread and all occur on other continents, thus providing some preliminary indication that the temporary wetland rotifers in the region comprise largely cosmopolitan species.

236 Tardigrada

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237 Phylum Tardigrada ('waterbears') are tiny coelomate animals (50 µm to 1.2 mm in length) 238 represented by approximately 900 species worldwide, of which only 62 species (representing 239 13 genera) are truly aquatic (Garey et al., 2008). Very little is known about the aquatic 240 tardigrades in the region and there has been no definitive study on the group. They are not 241 commonly encountered by freshwater ecologists and are easily overlooked due to their small 242 size (Rayner, 2002). No taxonomic or ecological information on the aquatic tardigrades of 243 southern Africa is available. There are however a number of studies on the terrestrial and 244 semi-terrestrial tardigrade fauna of mosses, lichens and leaf litter habitats in the region (for 245 reviews see Meyer & Hinton, 2009; Zawierucha et al., 2012). Despite the lack of studies, it is 246 strongly suspected that tardigrades inhabit southern African temporary wetlands, given their 247 exceptional adaptations to desiccation and physico-chemical extremes (reviewed by 248 Williams, 2006; Møbjerg et al., 2011). 249 Tardigrades are able to spend long periods in a desiccated cryptobiotic state, during which 250 time they are unique among metazoans in their ability to withstand extreme physico-chemical 251 fluctuations, tolerating laboratory immersion in liquid helium at -272°C and temperatures as 252 high as 340°C (Williams, 2006). During cryptobiosis, tardigrades are also easily dispersed by 253 wind, as are their eggs. Once their environment is re-flooded with water they rehydrate and 254 become active (Williams, 2006). Incredibly, tardigrades can survive for up to 70 years in the 255 cryptobiotic state (Rayner, 2002). Life spans are more commonly less than one year however 256 (Rayner, 2002). 257 Annelida 258 Annelid worms found in temporary wetlands in the region include earthworms (subclass 259 Oligochaeta) and leeches (subclass Hirudinea). Although sabellid polychaetes (class 260 Polychaeta) have been recorded elsewhere in temporary wetlands (Yozzo & Diaz, 1999), they 261 have not been reported in southern African. Ceratonereis keiskama Day, 1953 appears to be 262 the only polychaete worm recorded in entirely freshwater environments in SA, where it has 263 been recorded from permanent freshwater lakes that are estuarine relicts, being particularly 264 abundant in Lake Sibaya in KwaZulu-Natal (Day & Day, 2002). 265 Approximately one-third of the almost 5 000 valid species of oligochaetes worldwide are

aquatic, of which approximately 1 100 are found in freshwater environments (Martin et al.,

2008). About 50 microdrile and 20 megadrile taxa have been recorded from southern Africa (van Hoven & Day, 2002). Whilst most of the microdrile species are cosmopolitan, the megadriles are mostly African endemics (van Hoven & Day, 2002). None of the taxonomic works specifically report on which oligochaete taxa are found in southern African temporary wetlands, but the megadrile earthworm *Lumbriculus variegatus* (Müller, 1774) (cosmopolitan species) and microdriles belonging to *Nais* (Naididae) have been reported in faunal lists from ecological studies of temporary wetlands in the region (e.g. Bird & Day, 2016). Oligochaete specimens from southern African wetland studies are most commonly reported at the ordinal level as 'Oligochaeta' (e.g. Anusa et al., 2012; Ferreira et al., 2012). Oligochaetes can survive desiccation in temporary wetland environments by forming dehydration-resistant cysts, empirically documented for lumbriculids (Anlauf, 1990) and naidids (Montalto & Marchese, 2005).

Leeches are found in most freshwater habitats, although they are sparse in fast-flowing water or wetlands that are highly ephemeral (Oosthuizen & Siddall, 2002). Leeches are, however, well adapted to live in temporary waterbodies, mostly due to their ability to disperse via mobile vertebrates such as waterfowl (Davies et al., 1982; Williams, 2006) or attached to large invertebrates (Oosthuizen & Siddall, 2002). Some leeches are capable of surviving droughts as adults in mucus-lined cysts (Williams, 2006). The excellent dispersal ability of leeches results in their distributions in temporary wetlands in other regions of the world being well known (Williams, 2006). There are approximately 50 species of leeches in southern Africa (Oosthuizen & Siddall, 2002), but it is unknown how many can tolerate living in temporary wetlands. Bird & Day (2016) reported two species, the African leech Helobdella conifera (Moore, 1933) and the globally widespread H. stagnalis (Linnaeus, 1758) from temporary depression wetlands in Cape Town, SA. Oosthuizen & Siddall (2002) have recorded *Placobdelloides multistriata* (Jahansson, 1909) from temporary wetlands throughout southern Africa (the species is distributed across Africa), although they remark that the leeches usually enter small temporary pools inadvertently while attached to water scorpions and other hemipterans as transport hosts. All three of the above-mentioned species occurring in temporary wetlands thus have wide geographic distributions.

Mollusca

Approximately 111 species of gastropod and 26 bivalves have been reported from southern Africa, which includes 10 alien gastropod species (Appleton, 2002b). The region has very

299 low freshwater molluscan endemicity at about 12.3% (Appleton, 2002b). According to 300 Brown's (1994) habitat records, the following species are frequently found in temporary 301 wetlands: Pila occidentalis (Mousson, 1887); Lanistes ovum Troschel, 1845; Tomichia 302 ventricosa (Reeve, 1842); Ceratophallus natalensis (Krauss, 1848); Lymnaea natalensis 303 Krauss, 1848; L. columella (introduced from North America, now widespread in Africa); 304 *Physa acuta* Draparnaud, 1805 (introduced from Europe, now widespread in Africa); 305 Biomphalaria pfeifferi (Krauss, 1848); Bulinus globosus (Morelet, 1866) B. forskalii 306 (Ehrenberg, 1831); B. reticulatus (Mandahl-Barth, 1954); B. tropicus (Krauss, 1848); B. 307 scalaris (Dunker, 1845); and Aplexa marmorata (Guilding, 1828). Aplexa marmorata is an 308 invasive species from South America, which has recently been recorded from temporary 309 wetlands as far south as Cape Town by Mlambo et al. (2011). 310 The genus *Ferrissia* are freshwater limpets that are found in varied habitats including 311 seasonal pools throughout Africa (Brown, 1994). Although several studies have recorded 312 Ferrissia at the generic level in South African temporary wetlands (Mlambo et al., 2011; Bird 313 et al., 2013; Bird & Day, 2016), species level information is unavailable. Members of this 314 genus are able to partly close the shell using a septum, which facilitates aestivation during the 315 dry phase of a temporary wetland (Brown, 1994). 316 Of the African gastropods, the genus *Bulinus* excels in temporary wetland environments 317 (Vera et al., 1995). For example, B. depressus and B. scalaris were the species most 318 commonly found in isolated seasonal pools in Moremi Game Reserve, Botswana (Appleton 319 et al., 2003). Similarly, B. forskalii was the only mollusc collected together with 320 branchiopods in rain pools on the Makathini Flats (Hamer, 1989). The success of Bulinus 321 snails can, at least in part, be put down to their ability to aestivate for up to six months and 322 then exploit nutrient-rich temporary water bodies upon re-inundation by breeding rapidly (r-323 selected growth) during the short breeding season (Brown, 1994). 324 Bulinus and Biomphalaria are the two best-studied gastropod genera in southern Africa 325 because they are the aquatic intermediate hosts of human and bovine schistosomiasis (Rubaba 326 et al., 2016). Although species of *Biomphalaria* commonly occur in temporary habitats in the 327 Neotropical region (Paraense, 1996), this is rare for the African counterparts. *Biomphalaria* 328 species in the Neotropics often have lamellae in their shells' apertures that are thought to play 329 a role in tolerating desiccation. In a southern African context, only one specimen of B. 330 pfeifferi is known to have apertural lamellae (Appleton CC, personal communication).

Of the freshwater bivalves, members belonging to the family Sphaeriidae commonly inhabit temporary wetlands (Williams, 2006). The three genera of Sphaeriidae (Sphaerium, *Pisidium* and *Eupera*) present in southern Africa are in need of revision (Appleton, 2002b) and information on which sphaeriid species are known to inhabit temporary wetlands in the region appears to be lacking. Sphaeriids have been shown elsewhere to deal with desiccation by burrowing into the substrates of drying wetlands (McKee & Mackie, 1980). Most species require some residual moisture in the substrate to survive the dry season (at least 15% moisture for two Canadian sphaeriid species, McKee & Mackie, 1980) and thus are not likely to be encountered in highly ephemeral wetlands. Sphaeriids have effective dispersal mechanisms (e.g. clipping onto the limbs of mobile invertebrates and waterfowl) and are selffertilizing hermaphrodites, adaptations that further allow them to exploit temporary environments (Williams, 2006). Crustaceans: Malacostraca Most freshwater-dwelling members of the class Malacostraca (i.e. Amphipoda, Isopoda and Decapoda) lack physical adaptations for diapause or dispersal. Thus, they are not commonly associated with temporary wetlands but, due mostly to behavioural adaptations, certain species are able to persist in temporary waters (Williams, 2006). Amphipods appear to be very scarce in southern African temporary wetlands and virtually no records exist of their occurrence in these systems. Mlambo et al. (2011) and Bird & Day (2016) both recorded populations of the Gondwanan relictual species *Paramelita capensis* (Barnard 1916) and *P*. pinnicornis Stewart and Griffiths 1992 in small temporary wetlands of the Kenilworth Racecourse Conservation Area in Cape Town. However, studies of how these amphipods survive the summer dry phase have not been undertaken. A study of the North American amphipod Crangonyx pseudogracilis Bousfield, 1958 provides evidence that this epigean species is able to inhabit parts of the water table during the dry phase of a temporary wetland

and this mechanism explains both its ability to persist and its ubiquity across a network of

by epigean amphipods may be a key mechanism for amphipod survival in temporary

is reported from a congeneric North American amphipod, C. antennatus Packard, 1881,

nearby wetlands (Harris et al., 2002). The authors suggest that such subterranean explorations

wetlands in other parts of the world. Another interesting behavioural adaptation in amphipods

which survives the desiccated phase of mud-bottom pools by digging a burrow into the mud

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362 that is then blocked off from the outside with an air bubble (Holsinger & Dickson, 1977). The 363 burrow itself remains hydrated and provides a temporary aquatic refuge until the pool refills. 364 True crabs (Brachyura) also deal with desiccation by burrowing into the substrate. At least 365 four southern African species (namely Potamonautes lividus Gouws, Stewart and Reavell, 366 2001, P. flavusjo Daniels, Phiri and Bayliss, 2014, P. isimangaliso Peer & Gouws, 2015, and 367 P. calcaratus (Gordon, 1929)) exhibit a vaulted carapace, which is an adaptation to a semi-368 terrestrial lifestyle away from permanent waters (Daniels et al., 2014). All of these species 369 may appear superficially similar morphologically, but phylogenetically they are very distinct 370 (Daniels et al., 2014; Peer et al., 2015). From north-eastern KwaZulu-Natal, Potamonautes 371 lividus and P. isimangaliso are documented to live in close association with highly ephemeral 372 depression wetlands (Peer et al., 2015). These crabs occupy 0.3-0.5 m deep burrows within 373 the wetland substrate that remain moist throughout the dry season. Brachyurans associated 374 with temporary waterbodies worldwide have often also evolved physiological air-breathing 375 adaptations such as the ability to efficiently draw oxygen from moist air within burrows 376 (Cumberlidge, 2009), and this appears to be the case for *P. isimangaliso* (Peer et al., 2015). 377 Similarly, P. calcaratus displays the same ecological habits as P. isimangaliso, surviving in 378 burrows around ephemeral pools in the KNP (Daniels et al., 2014). Another congeneric, P. 379 flavusjo, burrows into peat soils in Highveld wetlands in Mpumalanga province where it 380 remains submerged in water by following the vertical movement of the water table (Daniels 381 et al., 2014). 382 The only freshwater isopods known to inhabit temporary wetlands in the region belong to 383 the genus Mesamphisopus (Gouws et al., 2005). Barnard (1926) observed that 384 Mesamphisopus abbreviatus (Barnard, 1927) burrows into muddy substrata in small 385 mountain streams that seasonally run dry. It aestivates in these burrows by lowering its

To our knowledge, and based on extensive literature searching, there are no records of prawns or shrimps in the freshwater temporary wetlands of southern Africa (indigenous freshwater crayfish do not occur in Africa).

province (Mlambo et al., 2011; Bird et al., 2013).

metabolism and breathing the moist air in the burrows, in a similar fashion to *P. isimangaliso*.

Mesamphisopus has been reported from temporary depression wetlands in the Western Cape

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392 Crustaceans: Large branchiopods

The class Branchiopoda, as a matter of convenience rather than taxonomic distinction, are usually treated as two groups: the large branchiopods (fairy shrimps, tadpole shrimps and clam shrimps) and the cladocerans (water fleas). Following suit, we address the cladocerans in a separate section together with their microcrustacean counterparts, the ostracods and copepods.

The large branchiopods are the enigmatic flagship invertebrates of temporary wetlands worldwide (Brendonck et al., 2008). They are specialists of lentic environments that incorporate a dry phase, although in some cases they do occur in fishless semi-permanent waterbodies (Ferreira et al., 2011). Their adaptations to ephemeral waterbodies have led to their prevalence in arid and semi-arid regions, where such waterbodies predominate. This is no exception in the largely semi-arid subcontinent of southern Africa, where they are widespread and diverse. The large branchiopod fauna is well studied in the region compared to most of the aquatic invertebrate groups so far covered in this review and to date approximately 66 species have been recorded (Tuytens et al., 2015).

Large branchiopods generally exhibit a lifestyle of rapid growth and maturation, followed by the production of drought-resistant eggs that can lie dormant in dry sediment under harsh conditions for decades and perhaps even centuries (Brendonck et al., 2008). Generally speaking, populations adopt a 'bet-hedging' strategy in that not all eggs hatch with the first onset of inundation, but rather some eggs remain dormant and only hatch after subsequent inundations so as to ensure survival of the population in a wetland that dries before a cohort reaches maturity (Simovich & Hathaway, 1997; Brendonck et al., 1998; Brendonck et al., 2008). Eggs are generally dispersed passively to other waterbodies via wind (Brendonck & Riddoch, 1999; Vanschoenwinkel et al., 2009), ingestion by predators, which transport the eggs in their gut (Rogers, 2014, 2015), or by attachment to mobile fauna (Vanschoenwinkel et al., 2011). More specific aspects of the adaptations of large branchiopods to temporary wetlands environments, including their biology and reproduction, are covered elsewhere (for reviews see Belk et al., 1991; Brendonck, 1996; Brendonck et al., 1998; Dumont & Negrea, 2002; Rogers, 2009).

Various aspects of the ecology of southern African large branchiopods (including metacommunity dynamics, competition and predation, adaptations to drying and local abiotic

423 conditions, human impacts, management and conservation) have received considerable 424 research attention due largely to a series of studies of rock pools in the eastern Free State 425 province (Vanschoenwinkel et al., 2007; Vanschoenwinkel et al., 2009; Vanschoenwinkel et 426 al., 2010a; Vanschoenwinkel et al., 2010b; Pinceel et al., 2013; Vanschoenwinkel et al., 427 2013; Tuytens et al., 2014) and south-eastern Botswana (Brendonck et al., 1998; Brendonck 428 et al., 2000a; Brendonck et al., 2000b; De Roeck et al., 2005; Jocqué et al., 2006; Jocqué et 429 al., 2010). Recently, the large branchiopod fauna of soft-bottomed wetlands and rock pools 430 have received ecological attention in Zimbabwe (Nhiwatiwa et al., 2011; Anusa et al., 2012; 431 Nhiwatiwa et al., 2014; Tuytens et al., 2015; Nhiwatiwa et al., 2017a; Nhiwatiwa & Dalu, 432 2017; Nhiwatiwa et al., 2017b) and the Western Cape province (De Roeck et al., 2007; De 433 Roeck et al., 2010; Mlambo et al., 2011). A scattering of studies exists from various other 434 localities, including Namibia (Day, 1990; Curtis, 1991), Botswana (Brendonck & Riddoch, 435 1997), the Drakensberg (Hamer & Martens, 1998), and soft-bottomed wetlands in the Free 436 State (Seaman et al., 1991; Meintjes, 1996), Mpumalanga (Ferreira et al., 2011, 2012), North-437 West (de Necker et al., 2016), Northern Cape (Hamer & Rayner, 1996) and KwaZulu-Natal 438 (Hamer & Appleton, 1991) provinces of SA. As far as we are aware, no ecological studies on 439 large branchiopods have been recorded from Mozambique. 440 The best-known of the large branchiopods is the order Anostraca (fairy shrimps). Early 441 taxonomic descriptions in the southern African region were pioneered by G.O. Sars and E. 442 Daday, but largest contribution to anostracans systematics was made by K.H. Barnard of the 443 South African Museum during the 1930s-1940s. During the 1990s, various workers updated 444 diversity and distribution data for the anostraca of the region. Hamer et al. (1994) and 445 Brendonck & Coomans (1994) reviewed the Streptocephalidae south of the Zambezi and 446 Kunene rivers and reported a total of 16 species. Brendonck (1995) added a new genus and 447 species Rhinobranchipus martensi Brendonck, 1995 (Branchipodidae) to the southern 448 African list of anostraca, which was collected from a shallow depression wetland in the 449 Thomas Baines Nature Reserve in the Eastern Cape province. Hamer (1999) provided a 450 taxonomic key for identification of the 46 southern African species of anostracans (six genera 451 and four families) known at the time, all of which are known to occur in temporary 452 freshwater wetlands with the exception of members of the genera Artemia and Parartemia, 453 which are adapted to hypersaline salt lakes and salt pans. Nhiwatiwa et al. (2017b) recently 454 described a new species, Stretocephalus sangoensis Nhiwatiwa, Dalu & Brendonck, 2017 in 455 the south-eastern Lowveld of Zimbabwe, which was formerly published as *Streptocephalus*

cf. *bidentatus*, indicating that more research is still required to prove valid species status and identify more taxa.

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Brendonck et al. (2000b) provided a focussed review on the genus *Branchipodopsis*, the members of which are shown to be specialists of transparent, very fresh (<50 µS cm⁻¹) and often highly ephemeral rock pools in the region. They described *Branchipodopsis* species as the 'record holders' of an ephemeral lifestyle in these rock pools, where populations are able to grow to maturity within the first week after inundation. Their long-distance dispersal ability by wind appears to be rare and rather they disperse to other nearby pools via overflows. This is suggested as a potential factor behind the high levels of endemism of *Branchipodopsis* species within the region. Since 2000, two new anostracan species have been described from KwaZulu-Natal province belonging to the genus *Metabranchipus* (*M. rubra* Rogers and Hamer, 2012 and *M. prodigiosus* Rogers and Hamer, 2012) of the family Branchipodidae.

Otherwise, the only recent notable taxonomic work in southern Africa concerning the anostracans is for the genus Artemia (Artemiidae, commonly known as brine shrimp). Artemia has been in a state of taxonomic confusion for quite some time (Hamer, 1999). The genus is cosmopolitan and restricted to salt lakes and salt pans with salinity levels above approximately 40 g L⁻¹, where fish and many predatory invertebrates are absent (Kaiser et al., 2006). In southern Africa, populations seem to be a mix of bisexual and parthenogenetic forms (Kaiser et al., 2006). The sexually reproducing species cannot be distinguished by gross morphological characters and require genetic analyses to separate species. Since such work had not been conducted in the region until recently, members of the genus have been referred to at generic level. However, Baxevanis et al. (2014) attempted the genetic identification of Artemia species in SA. Phylogenetic and network analyses revealed that three out of the seven studied populations belong to the invasive A. franciscana Kellog, 1906, while four belong to A. salina (Linnaeus, 1758). This is the first confirmed report of the invasive A. franciscana in the region and highlights concern regarding its invasive ability, as demonstrated by its complete replacement of the native A. salina population at the Veldrift Saltworks on the west coast of SA. Given that Baxevanis et al. (2014) only investigated seven populations, the taxonomic status of *Artemia* populations in the broader southern African region stills requires investigation.

Brendonck & Riddoch (1997) conducted a comprehensive survey of the anostracan fauna of Botswana, sampling from approximately 200 localities around the country, collecting both live specimens and mud samples containing eggs. They reported 14 anostracan species, with only one (Branchipodopsis kalaharensis Daday, 1910) being endemic to Botswana. They attribute the higher species richness (33) and endemicity (64%) in SA to greater habitable surface area, topographic and environmental heterogeneity, and research effort. Hamer & Brendonck (1997) expanded on the work of Brendonck & Riddoch (1997) by establishing a species-level checklist of the anostracan fauna for the whole region, providing a detailed distribution of each species. They divided the anostracan fauna into ten biogeographic categories, shaped largely by climatic factors: (i) widespread species occurring across southern Africa; (ii) eastern escarpment species occurring at high altitude within the Drakensberg; (iii) arid south-west species occurring in the arid and semi-arid regions of Namibia and the Karoo; (iv) tropical/subtropical species occurring on the low-lying eastern margin of southern Africa; (v) southern savannah species; (vi) Highveld species occurring in summer rainfall areas on the plateau at altitudes of 1000 – 1500 m; (vii) Cape east coast species occurring in winter-rainfall areas of the southern and south-eastern Cape margins; (viii) Cape west coast species occurring from Cape Town northwards along the west coast; (ix) Eastern Cape inland species (two species), of which Streptocephalus dregei Sars, 1899 occurs in the more arid lowland areas towards the south, whilst S. spinicaudatus Hamer and Appleton, 1993 occurs more northwards at higher altitude and with higher annual rainfall; and (x) Zimbabwean species that have been collected mainly from Zimbabwe and also Zambia, but not further south. The order Notostraca (tadpole shrimps) are branchiopod crustaceans that are known from as far back as the Triassic Period, earning them the status of 'living fossils' (Fryer, 1988; Vanschoenwinkel et al., 2012), although recently this concept for the group has been challenged (Mathers et al., 2013). Rayner (1999) provides an overview of the morphology, habitat preferences, feeding, reproduction and identification of southern African notostracans. Two species of *Triops* have been recorded from southern Africa, *T. granarius* (Lucas, 1864) and T. cancriformis (Bosc, 1801), although the latter is rare and has only been recorded from Ovamboland (northern Namibia) and the Kalahari (Hamer & Rayner, 1995). T. granarius is common and widespread in the drier areas of southern Africa (<500 mm mean annual rainfall), preferring muddy temporary pools seldom exceeding one hectare in size (for more

detailed distribution records see Hamer & Rayner, 1995). Triops taxonomy in southern Africa

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521 Hundsdoerfer (2006) used ribosomal DNA markers to investigate genetic distinctiveness of 522 T. granarius populations from Tunisia, Namibia and Japan, and their results strongly suggest 523 that the three populations belong to different, possibly cryptic species. Thus, further 524 molecular phylogenetic work on African *Triops* populations is required to more clearly 525 resolve cryptic species complexes. 526 The third major group of large branchiopods frequenting temporary wetlands are the clam 527 shrimps, although the colloquial term is used here solely for descriptive convenience. The 528 clam shrimps consist of the order Laevicaudata (smooth clam shrimp, one family with three 529 genera and 39 species) and, in the order Diplostraca, two suborders: Spinicaudata and 530 Cyclestherida, with approximately 116 known species (four families and 16 genera) 531 worldwide (Brendonck et al., 2008; Rogers et al., 2012). All three groups are primitive and 532 known as far back as the Devonian (Gueriau et al., 2016). Despite being abundant and 533 widespread in arid-zone temporary wetlands of the region (and worldwide), the taxonomy of 534 the clam shrimps has been poorly studied. The systematics has been problematic and 535 complicated by morphological plasticity in the fine details, despite generally uniform gross 536 morphology (Rogers et al., 2012). The difficulty in finding diagnostic morphological 537 characters appears to be a prime reason why virtually no taxonomists have tackled the 538 systematics of southern African clam shrimps since the studies of G.O. Sars, R. Gurney, G.S. Brady, E. Daday, K.H. Barnard and V. Brehm. 539 540 Brendonck (1999) provides a taxonomic key to distinguish the southern African species of 541 clam shrimps, although only genus-level distinction is provided in some cases (e.g. Cyzicus 542 Audouin, 1837), due to difficult taxonomy. Brendonck (1999) also provides distribution 543 records for the 18 species known at the time of his assessment. The only notable taxonomic 544 work that includes southern African clam shrimps in recent times is a revision of the 545 worldwide genera of the Limnadiidae (Spinicaudata) by Rogers et al. (2012), who combined 546 morphological and molecular information to show the presence of eight well-defined genera, 547 including two new genera (Afrolimnadia Rogers, Rabet & Weeks, 2012 and Calalimnadia 548 Rogers, Rabet & Weeks, 2012). South African material was used for the description of 549 Afrolimnadia alluaudi (Daday, 1926). Afrolimnadia was subsequently renamed 550 Gondwanalimnadia Rogers, Rabet & Weeks, 2016 because it is a junior homonym of

(and worldwide for that matter) is not without controversy. For instance, Korn &

551 Afrolimnadia Tasch, 1987, an fossil genus of spinicaudatan clam shrimps (Rogers et al. 552 2016). 553 Crustaceans: Ostracoda 554 Southern Africa has a diverse ostracod fauna. Martens (2001) reported about 155 species (ca. 555 55 genera) belonging to the class Ostracoda from the region, but remarked that dozens of new 556 species await description. Since this publication, several new species and genera have also 557 been added to the regional list (Martens, 2003, 2007; Jocqué et al., 2010), including the re-558 discovery of two species of giant ostracod (Liocypris grandis Sars, 1924 and Afrocypris 559 barnardi Sars, 1924) that were thought to be extinct (Martens, 2003; Matzke-Karasz & 560 Martens, 2007), both from temporary wetland habitats. The most striking feature of the 561 ostracod fauna of the region is the high endemicity of the Western and Eastern Cape 562 provinces, which although possessing few endemic genera, harbour many endemic species 563 (de Moor & Day, 2013). The northern and central parts of Namibia and Botswana and the 564 southern part of Zimbabwe form another belt of high endemism, with generic endemism in 565 these areas being comparable to the exceptionally rich and endemic ostracod fauna of Lake 566 Tanganyika (Jocqué et al., 2010). The ostracod fauna of southern Africa exhibits its highest 567 endemism and diversity in temporary wetland environments where few cosmopolitan species 568 occur (Martens, 2001). Ostracods are mostly benthic scavengers (although some forms are 569 pelagic) and play an important role in temporary wetland ecosystems as detritivores 570 (Martens, 2001). The success of ostracods in temporary wetlands is largely attributed to their 571 production of highly desiccation-resistant eggs, but also their ability to reproduce 572 parthenogenetically and the ease with which their eggs are passively dispersed to other 573 waterbodies via wind dispersal or attached to other animals (for review see Horne & Martens, 574 1998). 575 Martens (2001) provides a taxonomic key for the genera (and species for some recently-576 revised genera) of southern African freshwater ostracods, many of which are found in 577 temporary wetland environments. Following this reference, the genera that have been 578 unequivocally recorded from temporary wetlands are: Cypricercus, Eundacypris, 579 Gomphocythere, Globocypris, Korannacythere, Leucocythere, Megalocypris, 580 Ovambocythere, Physocypria, Pseudocypris, Ramotha, Sarscypridopsis, Sclerocypris, 581 Zonocypris. Jocqué et al. (2006) reported Potamocypris and Strandesia from temporary rock 582 pools in Botswana, while Amphibolocypris (A. arida and A. exigua) were recorded from

583 temporary rock pools in Botswana and a temporary soft-bottomed wetland in KNP, 584 respectively (Jocqué et al., 2010). 585 In the Western Cape province, Martens (2007) recorded the genus *Mnementh* from 586 temporary wetlands; and Martens et al. (1996) reported Zonocypris cordata Sars, 1924 and 587 Physocypria capensis Sars, 1896, which are both found in temporary and permanent 588 wetlands. Additionally, Bird & Day (2016) recorded the following genera *Chrissia*; 589 Sarscypridopsis; and Gomphocythere from temporary depression wetlands within the 590 Kenilworth Racecourse Conservation Area, Cape Town. This also included the following 591 species: Cypretta turgida Sars, 1896, Cypricercus episphaena (Müller, 1908), Cypridopsis 592 vidua (Müller, 1908), Paracypretta acanthifera Sars, 1924, P. capensis, Ramotha capensis 593 (Müller, 1908), and Zonocypris cordata Sars, 1924. 594 Reviewing the ostracod fauna of KwaZulu-Natal, Martens et al. (1998) report that most of 595 the diversity in lowland temporary wetlands of the province is in the genera *Heterocypris* and 596 Cypricercus, but diversity and endemicity in these habitats is generally low. The higher 597 altitude temporary rock pools of the Drakensberg have high diversity and endemicity in the 598 genera Sarscypridopsis and Heterocypris. However, most notable of the Drakensberg pools is 599 the diversity within the family Limnocytheridae that is unmatched anywhere else in the 600 world. 601 Crustaceans: Copepoda 602 Copepods are one of the most diverse crustacean groups, including over 14,000 known 603 species of both free-living and symbiotic forms. Most free-living copepods are marine, but 604 approximately 2,800 species have colonized a wide variety of freshwater habitats (Boxshall 605 & Defaye, 2008). Copepods in freshwaters represent a pivotal component of the food webs as 606 part of the zooplankton and benthic/littoral communities worldwide. Up to 20 genera and 51 607 species of freshwater copepods have been recorded from the region (Rayner, 2001). Most of 608 the information available for the region concerns the calanoid copepods (order Calanoida) 609 and there has been virtually no recent taxonomic research on the cyclopoids or harpacticoids. 610 Among the calanoids, only members of the exclusively freshwater family Diaptomidae have 611 been recorded in the region (Rayner & Heeg, 1994). The subfamily Diaptominae, containing 612 about 410 species, is represented in the region by the genera Tropodiaptomus and 613 Thermodiaptomus, both of which occur in more permanent waters and are not considered

614 further here. The diaptomid subfamily Paradiaptominae currently consists of 25 species 615 (Suárez-Morales et al., 2015), most of which are endemic to Africa and adapted to life in 616 temporary wetlands (Rayner, 2001). This largely endemic group includes a genus, Lovenula, 617 that comprises extremely large individuals (de Moor & Day, 2013). Rayner (2001) provides a 618 taxonomic key to the species of Diaptomidae in the region, along with cursory notes on the 619 cyclopoid and harpacticoid taxa that have been described or recorded from the region. The 620 following species in the subfamily Paradiaptominae have thus far been recorded from 621 temporary wetlands in the region (for distributional information, see Day, 1990; Rayner, 622 1998; Rayner, 2000, 2001; Suárez-Morales & Rayner, 2004; Suárez-Morales et al., 2015): 623 Lovenula falcifera (Lovén 1845), L. africana (Daday, 1908), L. simplex Kiefer, 1929, L. 624 raynerae Suárez-Morales, Wasserman & Dalu 2015, Paradiaptomus lamellatus Sars, 1895, 625 P. schultzei van Douwe, 1912, P. similis van Douwe, 1912, P. peninsularis Rayner, 1999, P. 626 hameri Rayner, 1999, Metadiaptomus capensis (Sars, 1907), M. purcelli (Sars, 1907), and M. 627 meridianus (van Douwe, 1912). 628 The cyclopoids (order Cyclopoida), on the other hand, are represented in freshwaters by 629 the widespread family Cyclopidae; in Africa this group is represented by 167 species, a figure 630 that resembles that of the Neotropical region (174 species), but is about half the number 631 found in the Palaearctic region (337 species) (Boxshall & Defaye, 2008). In southern Africa, records of cyclopids are scarce and mostly consist of cosmopolitan species. This is probably 632 the result of the 19th Century species concepts and delimitations that were extensively used 633 634 and applied by European researchers who named species from different geographic regions 635 on the basis of their resemblance with European species. Revisionary studies have proved 636 that records of some of these "cosmopolitan" nominal species (i.e. Acanthocyclops vernalis 637 (Fischer, 1853) and A. robustus (Sars, 1863)) represent species complexes containing 638 undescribed taxa. The South African cyclopoid copepod fauna certainly deserves further 639 examination with recent taxonomic standards. The following cyclopoids have been recorded 640 in temporary wetlands in the region (for distributional information, see Day, 1990; Rayner, 641 2001): A. vernalis, Mesocyclops major Sars, 1927, Thermocyclops oblongatus Sars, 1927, T. 642 macracanthus Kiefer, 1929, T. schuurmanae Kiefer, 1928, Microcyclops crassipes (Sars 643 1927), M. inopinatus (Sars, 1927), Eucyclops (Afrocyclops) gibsoni (Brady, 1904), E. 644 sublaevis (Sars, 1927), E. serrulatus (Fischer, 1851), Tropocyclops prasinus (Fischer, 1860), 645 Ectocyclops phaleratus (Koch, 1838), Paracyclops poppei (Rehberg, 1880), and P. 646 fimbriatus (Fischer, 1853).

Information on the African freshwater harpacticoids (order Harpacticoida) reveals a moderate diversity (108 species), which is comparable to that of India and Thailand. The most diverse harpacticoid family, the Canthocamptidae is represented by only 46 species in Africa, whereas more than 320 occur in the Palaearctic region (Boxshall & Defaye, 2008). Records of Harpacticoida in southern Africa are extremely scarce, let alone which taxa are known from temporary waters. *Nitokra dubia* Sars, 1927 and *Elaphoidella bidens coronata* Kiefer, 1934 have both been recorded from temporary wetlands of the Cape Flats near Cape Town, SA (Rayner, 2001). Vanschoenwinkel et al. (2008) recorded *Bryocamptus* sp. from temporary rock pools of the eastern Free State province. Rayner (2001) lists *Attheyella natalis* Brady, 1904, *A. warreni* Brady, 1913, *Harpacticus meridionalis* Sars, 1927, *Cletocamptus confluens meridionalis* Kiefer, 1934 and *C. trichotus* Kiefer, 1934 as other harpacticoid species that occur in southern Africa, but information could not be sourced on whether these species occur in temporary wetlands.

Of the ecological work directed specifically at copepods, most has focussed on aspects related to diapause. Diapause is a stop in the ontogenetic development induced by the need of physiological adjustments in the individual; in copepods this process affects one stage only, occurs regularly and is only partially related to environmental factors. Dormancy is a reversible cessation of development clearly related to environmental changes and involves different stages (depending on the species and environmental scenario) including eggs, nauplii, copepodites and adults (Dahms, 1995). Eggs are the most common dormant stage among diaptomids, and in cyclopoids dormancy is frequently related to copepodites C4 or C5 (Suárez-Morales et al., 2015). Diapause occurs under drying or unfavourable environmental conditions and is a dominant feature of the life cycle of many copepod species and a major factor in their successful colonisation of temporary wetland habitats worldwide (Dahms, 1995; Williams, 2006). In temporary ponds, cyclopine cyclopoids can emerge from diapause within a single day of the appearance of water (Reid & Williamson, 2010). Unlike ostracods, copepods generally reproduce sexually, but often have very fast development times, as little as one week to reach the adult stage, depending on prevailing environmental conditions (Williams, 2006). Similarly to ostracods, however, eggs are passively dispersed via wind or attached to mobile fauna (Dahms, 1995). Very little other ecological work has been directed specifically at copepods, with the exception of Wasserman et al. (2016a), Dalu et al. (2017b), Dalu et al. (2017c) and Wasserman et al. (2018), who all investigated aspects of the trophic

679 ecology of the paradiaptominids L. raynerae and P. lamellatus in the Eastern Cape province of SA. 680 681 Crustaceans: Cladocera 682 In contrast to copepods, most of the world's over 600 species (Forró et al., 2008) of 683 Cladocera ('water fleas') occur in freshwater environments, where they often dominate the 684 lentic fauna in terms of abundance. Southern Africa harbours a mixture of Afrotropical, 685 Palaearctic, circumtropical and local endemic elements in its cladoceran fauna (Smirnov, 686 2008; Van Damme et al., 2013). The taxonomy of cladocerans in the region requires an 687 urgent overhaul, particularly the Chydoridae and the Daphniidae (Seaman et al., 1999; Van 688 Damme et al., 2013), thus an estimate of the number of species in the region is not presently 689 available. Smirnov (2008) reported 112 cladoceran species from SA, with a wide 690 representation of Palearctic species. Traditionally, the region was considered to possess low 691 levels of cladoceran endemism (Frey, 1993; Korovchinsky, 2006; Smirnov, 2008), but more 692 recently this has been attributed to the limited state of knowledge of the fauna rather than 693 genuinely low endemism (Van Damme et al., 2013). As is the pattern for Ostracoda and 694 Copepoda, the greatest cladoceran species richness and endemicity in the region occurs in 695 temporary wetland environments, particularly montane rock pools in the east (e.g. 696 Drakensberg) and endemics of Western Cape lowland pools (Van Damme et al., 2013). The 697 families Chydoridae and Macrothricidae, the sub-genus Daphnia (Ctenodaphnia) and the 698 genera Ceriodaphnia and Moina are common inhabitants of temporary wetlands in the region 699 (Seaman et al., 1999). A literature search of taxonomic and ecological records of cladocerans 700 from temporary wetlands in the region was attempted, but given the urgent need of 701 taxonomic revision of species in this region (Van Damme et al., 2013), it was later 702 abandoned. 703 Survival strategies of cladocerans in temporary wetlands are broadly similar to those of the 704 other microcrustaceans, with highly desiccation-resistant eggs being produced as water levels 705 draw down (Vandekerkhove et al., 2005). Cladocerans favour parthenogenetic (asexual) 706 cloning during favourable times when the wetland is inundated and males are rare for much 707 of the hydroperiod (Williams, 2006). Males appear more frequently towards the end of the

hydroperiod and sexual reproduction prevails as water disappears from the wetland. Whilst

parthenogenetic cloning produces soft eggs for rapid development, sexually produced eggs

are encased in a hardened ephippium and can withstand drying, freezing and passive dispersal

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- to other waterbodies (Bilton et al., 2001; Vandekerkhove et al., 2005; Williams, 2006). As
- has been found in many aquatic environments globally, cladocerans have also been identified
- as important trophic components in systems of the region (Dalu et al., 2016; Wasserman et
- 714 al., 2016b; Dalu et al., 2017b; Dalu et al., 2017c).
- 715 Hydrachnidia: Water mites
- Water mites (suborder Hydrachnidia) are abundant and ubiquitous in temporary wetlands and
- over 6 000 species have been described worldwide from freshwater environments, including
- 718 160 species from southern Africa (Di Sabatino et al., 2008). Since then, more species have
- been added to the southern African records for Torrenticolidae (Goldschmidt & Smit, 2009),
- Hygrobatidae (Pešić et al., 2013), Mideopsidae (Pešić et al., 2013), Arrenuridae (Smit, 2012)
- and Hydryphantidae (Gerecke, 2004). Jansen van Rensburg (1976) provided a taxonomic key
- to identify the 22 water mite families from the region (updated by Jansen van Rensburg &
- Day, 2002). Throughout the key, and indeed also for more recent species-level accounts (see
- above), no mention is made in the habitat descriptions for each taxon as to the preference for
- temporary versus permanent water bodies. Hence, we are unable to provide further
- information from the taxonomic literature regarding those taxa specifically known to inhabit
- temporary wetlands in the region. The following taxa have been recorded from temporary
- wetlands in the south-western Cape Mediterranean-climate region of SA (records from
- Mlambo et al., 2011 unless otherwise specified), with the help of Dr R. Gerecke, Tübingen,
- Germany, and Dr E. Ueckerman, ARC, Pretoria: *Arrenurus* spp. (Arrenuridae), *Eylais* spp.
- 731 (Eylaidae), Hydrachna fissigera Koenike, 1898, Hydryphantes parmulatus Koenike, 1912,
- 732 Diplodontus schaubi (Koenike, 1893), Mamersa testudinata Koenike, 1898, Limnochares
- 733 crinita Koenicke, 1898, Piona sp. (Pionidae), and Neumania spp. (Unionicolidae). In
- addition, hygrophilous representatives of several terrestrial mite taxa were reported by Bird et
- al. (2013) from the cohorts Gamasina (Macrocheles sp., Macrochelidae), Astigmatina
- 736 (Oribatida), and Parasitengona (Trombidiidae).
- The great diversity of water mites in temporary habitats is remarkable, given that most do
- not possess the typical adaptations of temporary wetland invertebrates (either diapause of
- eggs/adults or active dispersal as adults) (Bohonak et al., 2004). Instead, the radiation of
- vater mites in temporary wetlands has been facilitated by parasitic larvae that, once hatched
- from eggs laid within a waterbody, parasitize adult insects and are thus dispersed to other
- 742 waterbodies (Bilton et al., 2001; Bohonak et al., 2004). Parasitism is obligate in all species

743 studied so far that colonize a tatic water bodies, and mite larvae die if they fail to penetrate an 744 insect host (Bohonak et al., 2004). However, not all water mites survive the dry phase via 745 parasitism, and certain taxa (e.g. some members of the Hydryphantidae) are capable of 746 surviving dry phases as deutonymphs or adults that enter an inactive state in microhabitats 747 with persistent moisture, such as moist mud or damp crevices in logs (Wiggins et al., 1980; 748 Bohonak et al., 2004). 749 *Insecta: Ephemeroptera (mayflies)* 750 The order Ephemeroptera have the distinction of being the most ancient lineage of extant 751 winged insects, dating back to the late Carboniferous or early Permian periods (Brittain & 752 Sartori, 2003; Barber-James et al., 2008). Despite over 3 500 species worldwide and 435 753 species from the Afrotropical region (Sartori & Barber-James, 2018), most mayfly diversity 754 is represented in running waters and this ancient group is relatively depauperate in lentic 755 environments (Newell & Hossack, 2009). A baetid species, Cloeodes hydation McCafferty & 756 Lugo-Ortiz, 1995, from temporary wetlands (rock pools, ephemeral pools and springs) of the 757 Pantanal wetland system was the first mayfly species to be experimentally shown to 758 withstand repeated drying conditions of up to nine hours at a time (Nolte et al., 1996). On the 759 contrary, a known inhabitant of intermittent waters, *Daleatidium* spp. (Leptophlebiidae), from 760 New Zealand dies only after three hours of drying exposure (Robinson & Buser, 2007). 761 However, Nolte et al. (1996) made a distinction between C. hydation and the chironomid 762 Polypedilum vanderplanki Hinton, 1951, calling it drying-tolerant as opposed to drying-763 resistant, which the latter is. Although the genus *Cloeodes* occurs in the region, a similar 764 characteristic of drying-tolerance has not been demonstrated, and it has also not been reported 765 from temporary wetlands. In southern Africa, only members of the genera Cloeon Leach, 766 1815 and *Procloeon* Bengtsson, 1915 (Baetidae) are known from truly lentic waterbodies 767 such as dams and wetlands and appear to be capable of reaching high abundance in 768 temporary waterbodies (Barber-James & Lugo-Ortiz, 2003). Similar results have been 769 reported in Irish turloughs (Reynolds, 2000). 770 Although the aquatic nymphal phase of these baetid mayflies is rather delicate and 771 desiccation-intolerant, populations persist in temporary wetlands due to the nymphs being 772 extremely fast-growing (reaching maturity in 2-3 weeks) and the adults apparently producing 773 desiccation-resistant eggs (Williams, 2006). Dispersal is via short-lived winged adults or via

passive dispersal of eggs (Bilton et al., 2001; Williams, 2006). Barber-James & Lugo-Ortiz

775 (2003) list 22 Afrotropical species of *Cloeon* (nine from SA) and three species of *Procloeon* 776 (one from SA). It is not yet known which of these species inhabit temporary wetland 777 environments, which reflects the desperate need for more species-level studies of mayflies in 778 Africa (Barber-James & Gattolliat, 2012). Cloeon is widespread in the region and has been 779 recorded from temporary wetlands throughout the region, including the Western Cape 780 (Mlambo et al., 2011; Bird et al., 2013) and Mpumalanga (Foster et al., 2015) provinces of 781 SA, Namibia (Day, 1990), Zimbabwe (Chakona et al., 2008) and Botswana (Jocqué et al., 782 2006). 783 Gillies (1990) reported the unusual habitat of the baetid Acanthiops erepens (Gillies, 784 1990), which occurs in Tanzania on rock in semi-permanent vertical trickles and in the splash 785 zone next to small cascades. The nymphs cling to moss growing on the rock face that is kept 786 moist by spray from the adjacent waterfall, and if disturbed they plummet into the waterfall 787 pool below. 788 Insecta: Odonata 789 The order Odonata, comprising some 5,680 species worldwide (Kalkman et al., 2008), is 790 divided into the suborders Zygoptera (damselflies) and Epiprocta, which is further divided 791 into the infraorders Anisoptera (dragonflies) and Aniszygoptera (Asian species only) (Rehn, 792 2003). As with the Ephemeroptera, the odonates are an ancient order, dating back well into 793 the Permian (Kalkman et al., 2008). The Odonata are relatively well studied, especially in the 794 Afrotropical region where their diversity is depauperate compared to other tropical regions 795 (Dijkstra, 2003) and thus their taxonomy is relatively well known in comparison to other 796 freshwater invertebrate groups (but still not without some confusion, see Dijkstra, 2003). 797 That said, Dijkstra et al. (2015) recently described a further 60 new African species as adults, 798 from regions north of the area under review in this paper. Therefore, if brightly coloured, 799 charismatic creatures such as adult Odonata are still incompletely known, how much less do we yet know about the more obscure freshwater invertebrates, especially those inhabiting 800 801 temporary waterbodies in Africa. 802 Unlike many of the African freshwater invertebrates, IUCN Red List information has been 803 compiled for many of the southern African odonate species, with most of the threatened 804 species being habitat specialists occurring in highland areas of the Western and Eastern Cape 805 provinces (Samways, 2004), many frequenting temporary water bodies, but most as

opportunistic visitors rather than being dependent upon these habitats. Odonate species 807 exploiting temporary habitats depend on dispersal for survival and tend to have large home 808 ranges and are habitat generalists, being tolerant of ecological change (Clausnitzer et al., 809 2012). Thus, temporary wetland species are generally not threatened. In southern African arid 810 regions, no desert endemics are known and most of the widespread desert-inhabiting species 811 are powerful fliers belonging to the Anisoptera that are not restricted to deserts, being more 812 common in savannah wetlands (Suhling et al., 2003). The 889 known odonate species of the 813 Afrotropical region (Kalkman et al., 2008) all have aquatic larvae, which are not capable of 814 withstanding desiccation themselves (for a rare exception from Brazil, see Van Damme & 815 Dumont, 1999). Odonate populations are however, able to persist in temporary habitats 816 through production of drought-resistant eggs combined with fast-growing larvae, multivoltine 817 life cycles and strong active dispersal of adults (for more detailed reviews of odonate 818 adaptations to temporary habitats, see Johansson & Suhling, 2004; Suhling et al., 2005; De 819 Block et al., 2008; Hassall & Thompson, 2008). Timing of hatching may also be important. 820 Schiel & Buchwald (2015) showed that for 15 species of odonates across three genera (Lestes 821 - Lestidae, Aeshna - Aeshnidae and Sympetrum - Libellulidae), species inhabiting only 822 temporary ponds hatched significantly earlier in the year than congeners inhabiting a 823 combination of permanent and temporary ponds. Despite the harsh environmental conditions 824 often presented in temporary waterbodies, a major reward for exploiting such habitats is often 825 the reduced predation pressure on larvae (lack of fish for instance) allowing them to forage 826 actively for food, whereas in permanent waters larvae generally need to remain concealed and 827 rely on ambush predation (Hassall & Thompson, 2008). 828 Several checklists are available covering various sub-regions of southern Africa, from 829 which habitat preferences of the various species can be discerned (if such information is 830 available at all). Martens et al. (2003) listed 102 odonate species from Namibia. They listed 831 the following taxa as facultative inhabitants of temporary wetlands: Lestes pallidus Rambur, 832 1842, Ischnura senegalensis (Rambur, 1842), Anax ephippiger (Burmeister, 1839), Pantala 833 flavescens (Fabricius, 1798), and Sympetrum fonscolombii (Selys, 1840). Suhling et al. 834 (2003), in their investigation of desert-adapted odonates, listed the following species from 835 ephemeral pools in the Namib desert of Namibia: Africallagma glaucum (Burmeister, 1839), 836 Ischnura senegalensis (Rambur, 1842), Anax ephippiger, A. imperator Leach, 1815, 837 Paragomphus genei (Selys, 1841), Crocothemis erythraea (Brullé, 1832), Orthetrum 838 chrysostigma (Burmeister, 1839), Palpopleura lucia (Drury, 1773), P. flavescens, S.

839 fonscolombii, Trithemis annulata (Palisot de Beauvois, 1807) and T. kirbyi ardens 840 (Gerstäcker, 1891). Mlambo et al. (2011) recorded the anisopteran genera *Anax* (Aeshnidae) 841 and Trithemis (Libellulidae) and the zygopteran genera Africallagma, Ischnura and 842 Pseudagrion (all Coenagrionidae) from temporary wetlands of the Western Cape. Jocqué et 843 al. (2006) reported the larvae of *P. flavescens* as top predators in temporary rock pools of 844 south-eastern Botswana. Simaika et al. (2016) investigated the benefits of artificial ponds as 845 biodiversity refugia for Western Cape dragonflies and listed the following taxa from a 846 temporary pond: A. imperator, C. erythraea, I. senagalensis, Orthetrum chrysostigma 847 (Burmeister, 1839), O. trinacria (Selys, 1841) and Trithemis arteriosa (Burmeister, 1839). 848 Samways (1999) noted L. pallidus as a frequent inhabitant of temporary wetlands in SA, 849 while Clark & Samways (1996) recorded *Palpopleura deceptor* (Calvert, 1899), *P. jucunda* 850 Rambur, 1842, P. lucia, Bradinopyga cornuta Ris, 1911, T. kirbyi ardens and P. flavescens 851 from ephemeral rock pools in KNP. Interestingly, larvae of the libellulid *Hemistigma* 852 albipunctum (Rambur, 1842), which inhabits marshes and swamps, can remain alive in the 853 mud as these habitats dry out, surviving with minimal water and emerging as an adult before 854 the local area completely dries out (Samways, 2008). 855 Kipping (2010) provided a checklist of the odonates of Botswana and noted that most of 856 the country's odonates are widespread African savannah species that, although having a 857 preference for more stable habitats, are able to disperse over wide distances and breed in 858 temporary waterbodies in arid areas such as the Kalahari Desert. Suhling et al. (2009) 859 reported that true obligate migrant species of the arid Kalahari region include *P. flavescens*, 860 S. fonscolombii and A. ephippiger. Kipping (2010) recorded the following species as 861 facultative inhabitants of temporary wetlands of Botswana: Lestes dissimulans Fraser, 1955, 862 L. pallidus, L. pinheyi Fraser, 1955, A. glaucum, Azuragrion nigridorsum (Selys, 1876), 863 Ceriagrion suave Ris, 1921, I. senegalensis, A. ephippiger, A. tristis Hagen, 1867, 864 Brachythemis leucosticta (Burmeister, 1839), B. cornuta, Diplacodes luminans (Karsch, 865 1893), Orthetrum brachiale (Palisot de Beauvois, 1817), O. caffrum caffrum (Burmeister, 866 1839), O. chrysostigma chrysostigma (Burmeister, 1839), O. trinacria, P. deceptor, P. 867 flavescens, Rhyothemis semihyalina (Desjardins, 1832), S. fonscolombii, Tramea basilaris (Palisot de Beauvois, 1807), T. kirby ardens and Urothemis assignata (Selys, 1872). For 868 869 more detailed information on the distribution, habitat and Red List status of these species, the 870 reader is referred to Kipping (2010) and Samways & Simaika (2016).

In some odonate species there are temporal changes in geographic range in accordance with the wet and dry cycles of the El Niño Southern Oscillation. An extreme case was for *Aciagrion dondoense* Dijkstra, 2007, which was not known in SA prior to the year 2000 when it appeared in great numbers on the KwaZulu-Natal coast, having come down the coast with flooding farther north. A few years later it was gone from the country with the dry phase appearing (Samways, 2010). Several South African species are adapted to these cycles, with species like *Orthetrum robustum* Balinsky, 1965 retreating from temporary pools in the dry phase to permanent lakes, to return later to the temporary pools in the wet cycle (Samways, 2008). One last factor to consider is that rivers that stop flowing and have only pools ('kuile') in the dry season. Such pools in these rivers may attract species temporarily until the river flows again. There may even be a succession of species as the pools gradually dry out, with *Pseudagrion commoniae* (Förster, 1902) and *P. salisburyense* Ris, 1921 being the last to survive before the pools finally dry out (Samways, 2008).

Insecta: Hemiptera

Sternorrhyncha (aphids, whiteflies, and scale insects); Auchenorrhyncha (cicadas and hoppers); Coleorrhyncha (moss bugs); and Heteroptera (true bugs) (Capinera, 2008; Forero, 2008). It comprises some 82,000 described species, and possibly almost 200,000 species in total worldwide. The bugs associated with water all belong to the Heteroptera, particularly the semi-aquatic Gerromorpha and the predominantly aquatic Nepomorpha. Gerromorphs have hydrophobic tarsi and gracile bodies that allow them to stand on the water surface supported by the surface tension of the meniscus, while nepomorphs generally crawl or swim

underwater and typically have reduced antennae and ocelli. The Nepomorpha have a fossil

record dating back about 250 million years (Grimaldi & Engel, 2005).

The hemimetabolous insect order Hemiptera is usually considered to contain four suborders:

Most species have winged adults that disperse well and can therefore colonise temporary wetlands quickly. The nepomorph families all breathe air through plastrons or respiratory tubes, and the gerromorph families are epipleustonic, living just above the water surface rather than in the water, so dissolved oxygen does not limit the potential of a water body for colonisation by water bugs. The species in most families are generalist predators, and can therefore inhabit a wide variety of waters with flow rates that are slow or absent. Many gerromorphs associate with floating or emergent vegetation that helps them to avoid predators and being washed away in flowing waters (Reavell, 2003). Immature nymphs have

biologies similar to their adults, but cannot fly, so they complete their development only in temporary wetlands that take more than (generally) four to ten weeks to dry up. Adults of some species of gerromorph may be winged or apterous (regulated by temperature and photoperiod); when apterous adults inhabit a temporary wetland, they indicate that at least one generation has completed its life cycle in the wetland. The high mobility and broader habitat preferences of most aquatic bugs means that populations of temporary wetland species are generally not threatened (Collinson et al., 1995).

The morphology, taxonomy and general ecology of the southern African water bug fauna was summarised by Reavell (2003), who stated that little work had been done on the fauna of the region, and that it needed urgent taxonomic revision. Reavell (2003) provided a key to the families of aquatic Hemiptera in the region, and a list of the known genera, but keys to the genera and species are still lacking. Ecological and life history studies of temporary wetland water bugs in the region include Hutchinson (1933), Hynes (1955), Weir (1966), Tawfik (1969), Boettger (1974), Mlambo et al. (2011), Ferreira et al. (2012), Bird et al. (2013) and Nhiwatiwa et al. (2017a). Hemiptera from six families constituted an average of 14% of the individual invertebrates surveyed in a large number of temporary waters in the Western Cape (Mlambo et al., 2011), and Hemiptera were co-dominant with Coleoptera (beetles) in nine temporary pans in Mpumalanga (Ferreira et al., 2012). The dominant families are usually Corixidae and Notonectidae. The water bugs tend to be associated with warmer, alkaline water in temporary pans (Ferreira et al., 2012), and larger-bodied species are more common in the absence of fishes in ephemeral pans (Nhiwatiwa et al., 2017a; Nhiwatiwa et al., 2017b). However, because of their high vagility and relatively broad habitat tolerances, water bugs may be poor indicators of anthropogenic effects on temporary wetlands in SA (cf. Bird et al., 2013), an effect that can be compounded by artefacts of the sampling method (de Klerk & Wepener, 2013). Their instantaneous diversity may also be related to the stage of the wetland's hydroperiod (Lahr et al., 1999; Dalu et al., 2017b; Dalu et al., 2017c), since older wetlands have had more time to become colonised and develop prey populations, thus obscuring the effects of human activities.

Insecta: Trichoptera

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Trichoptera are holometabolous insects, mostly with an aquatic larval and a terrestrial adult phase. They have been recorded worldwide with the exception of some oceanic islands and the polar regions (de Moor & Scott, 2003). The Trichoptera World Checklist (Morse, 2011)

lists 14 548 species of 616 genera in 49 families recorded globally thus far. Of these, 1 193 species (8.2%) in 83 genera from 21 families are known to occur in southern Africa (Tobias & Tobias, 2008). Recently, some species were collected in temporary wetlands, despite the earlier assertion of Scott (1970) that Trichoptera do not inhabit temporary lentic waters of the region: Oxyethria velocipes (Barnard, 1934) (Jones, 2002; Day et al., 2010) and Athripsodes sp. (Bird, 2012), both from the Western Cape province; and *Ecnomus thomasseti* Mosely, 1932, *Oecetis* sp. and *Oxyethira* sp. from Mpumalanga province (Ferreira et al., 2012). Trichoptera found in temporary waters show several adaptations to this habitat in different life stages, but little is known about this in southern Africa as research has mainly been conducted on species from other regions, discussed below. Larvae use silk to construct transportable or fixed cases from material readily available, such as sand, small stones, sticks, leaves and algae. The ability to build cases has allowed trichopteran larvae to adapt to a wide range of environmental conditions: spring sources, mountain streams, splash zones of waterfalls, large rivers, lakes and temporary wetlands (de Moor & Scott, 2003). Many species are represented in cooler and faster-moving waters as dissolved oxygen is a limiting factor for habitat colonisation (de Moor & Scott, 2003).

Trichopteran larvae are apneustic, which means that they lack spiracles; they take up dissolved oxygen through the thin abdominal membrane by direct diffusion. Their abdominal surfaces are often expanded with tubular or branched gills that increase the surface area of the abdominal membrane. To ensure sufficient uptake of dissolved oxygen, larvae can create a current through their cases by abdominal undulation. This aids gas exchange when oxygen concentrations are low, as is the case in temporary wetlands (Williams, 1987; de Moor & Scott, 2003).

As water levels decrease during drying, faunal density increases and cases offer a protective body covering and camouflage to protect larvae from predation. A combination of case material (mineral rather than organic), shape (short, wide case with "hedgehog" structure rather than long tubular cases or flat purses) and defensive behaviour (retreating deep into the case and remaining motionless) are adaptive strategies employed by *Limnephilus externus* Hagen, 1861 against predation by diving beetle larvae (*Dytiscus* sp.) (Wissinger et al., 2006). Another species, *L. picturatus* McLachlan, 1875, evades predation of pupae by burrowing into soft substrates to pupate, whereas larvae of *Asynarchus nigriculus* (Limnephilidae) (Banks, 1908) build stronger mineral cases prior to pupation. Furthermore, both species have

faster developmental times, which reduce temporal overlap with predators (Wissinger et al., 2003). Pupae can develop while the water column has dried up and the soil is still moist (Wiggins, 1973). The case material is important with regard to pond-drying. Larvae of *L. coenosus* Curtis, 1834 construct cases using organic material whereas *L. vittatus* (Fabricius, 1798) larvae use mineral material. Organic material has been shown to retain moisture better than mineral material, preventing drying of larvae (Zamora-Muñoz & Svensson, 1996).

Cases also protect caddisfly larvae from inter- and intraspecific aggression as water levels decrease (Wissinger et al., 2004b). Aggressive behaviour includes fighting (foreleg wrestling, biting, case shaking) and mobbing of individuals (involving 5-10 larvae) which often results in death and cannibalism of conspecifics (Lund et al., 2016). Wissinger et al. (2004b) have shown that cases reduced or prevented cannibalism significantly among larvae of *A. nigriculus*. Cases also reduced vulnerability of three *Limnephilus* species (*L. externus*, *L. picturatus*, *L. secludens* Banks, 1914) to intraguild predation by *A. nigriculus* (Banks, 1908). Aggression among *A. nigriculus* in mesocosm experiments was higher in high-density treatment and even higher in the absence of dietary protein supplement (Lund et al., 2016). Although aggressive behaviour has trade-offs, such as injury and subsequently becoming a secondary victim, cannibalism provides an important protein- or lipid-rich dietary supplement that offers advantages later in the life-cycle.

Dietary experiments on *L. coenotus* and *L. vittatus* supplementing detritus diet with protein have shown that larger larvae survive drying better as they contain more water compared to smaller ones (Zamora-Muñoz & Svensson, 1996). Furthermore, body mass of adult *L. externus* significantly increased when ambient food treatment was supplemented with a protein-rich supplement (Jannot et al., 2008). Larval diet can have a significant long-term effect on adult fitness (Jannot, 2009) as female body size is correlated with fecundity and male body size with mating success (Wissinger et al., 2004a).

Adults of *L. externus*, *L. picturatus* and *A. nigriculus* emerge prior to pond-drying and females enter an ovarian diapause after mating (Wissinger et al., 2003). Summer diapause is an important adaptation to survive dry periods (Wiggins, 1973; Wiggins et al., 1980), which also protects eggs from predation, drying and freezing (Wissinger et al., 2003). Oviposition in temporary pools usually occurs above or near water, not directly in water as is normal for most trichopteran species (Wiggins, 1973). Eggs are deposited in a gelatinous egg-matrix, which is highly resistant to extremes like desiccation and freezing (Wiggins, 1973). Eggs are

deposited under rocks and logs, where larvae hatch and develop further within a gelatinous mass until re-wetting occurs (Wissinger et al., 2003). *Lectrides varians* Mosely, 1953 show a bimodal response – some larvae become active with the first onset of re-wetting or reimmersion whereas others remain aestivating and only become active when surface water levels are more stable. Although this adaptation ensures population survival, it is at the cost of larval survivorship (Wickson et al., 2012). Larvae enter the temporary ponds with, for example, melt water, develop further and pupate prior to pond-drying (Wissinger et al., 2003).

1007 Insecta: Diptera

Virtually all African temporary waters support the larvae of at least one species of fly, and flies boast of a large array of adaptive mechanisms for this variable environment (McLachlan & Ladle, 2001). In the region, the families Chironomidae, Ceratopogonidae and Culicidae commonly occur even in the Kalahari and Namib deserts, where rain may fall only at intervals of several years. Although dipterans are ubiquitous in these systems, very few species are able to survive desiccation (Williams, 2006). Unlike many branchiopod crustaceans, which withstand dry periods as desiccation-resistant cysts, most flies that occur in temporary waters survive the dry period as adults or, in the case of some culicids, as larvae in perennial water bodies. Hinton (1953) indicated that some African Stratiomyidae can withstand desiccation, though this is not recorded in southern Africa.

Chironomids mostly survive dry periods as adults. Frouz et al. (2003) reviewed the strategies employed by chironomids to survive the variable environment of temporary wetlands. Given that larvae of several species are found in hyper-arid conditions in the Namib Desert, where pools may not form for 5 to 10 years at a stretch, the question yet to be answered is, "How are the *adults* able to survive the dry years in these environments?" A few other species survive for some time in stout pupal cocoons during dry periods (Armitage, 1995), but presumably these periods are relatively short. When it comes to survival under harsh conditions, larvae of the 'sleeping chironomid', *Polypedilum vanderplanki* Hinton, 1951, hold the record. The species breeds in small unshaded rock pools in Central and West Africa. Larvae can undergo up to 17 years of desiccation and, when wetted, will 'wake up' and behave normally, feeding and growing until the next time they are desiccated. Hinton (1960) performed various experiments on these remarkable organisms, showing that they can withstand temperatures between -270°C and +102°C and can survive through a

wetting/drying cycle at least ten times. An additional species of 'sleeping chironomid', *P. ovahimba* Cranston, 2014, has been recently described from rock pools of the Waterberg Plateau, Namibia (Cranston, 2014).

Other chironomids, such as *Chironomus pulcher* Wiedemann, 1830, also breed in rain pools on rocks throughout much of Africa and have rapid life cycles. Larvae of the chironomid subfamily Podonominae (*Archaeochlus* sp. and *Afrochlus harrisoni* Freeman, 1964) are found in temporary trickles over rocks and boulders in the Drakensberg, on granite inselbergs near Harare, and in Namibia (Harrison, 2002). Similarly, *Harrisonina petricola* Freeman, 1956 was originally found breeding in the thin film of water running over stones in an ephemeral stream in the Olifants River Valley. It is not known how these survive dry periods. Rock pools in Africa are dominated by dipteran larvae (up to 300 000 per pool), which are often the only macroinvertebrate inhabitants. If the pools are very short-lived (1-2 days), the only inhabitants are larvae of *P. vanderplanki*. Pools lasting several weeks often support *Chironomus imicola* Kieffer, 1913, which has little tolerance of desiccation, but is able to complete its life cycle in 10-12 days (McLachlan & Cantrell, 1980).

Larvae of a few ceratopogonid species are found in extremely arid regions (de Meillon & Wirth, 2002) and yet they seem not to be particularly adapted to withstanding desiccation. There are no African examples. Larvae of *Culicoides sonorensis* Wirth & Jones, 1957 from the Sonoran Desert in North America, for instance, are able to survive drying, but only for a day or two (McDermott & Mullens, 2014). Pools lasting several days often contain larvae of the ceratopogonid *Dasyhelea thompsoni* de Meillon, 1936 that burrow into mud at the bottom of the pond when water evaporates (McLachlan & Cantrell, 1980). Similarly, Dodson (1987) found that *Dasyhelea subletti* Wirth, 1987 was the most common taxon in short-lived ephemeral rock pools in the Moab desert, USA.

Numerous species of culicid are associated with temporary waters in Africa (Coetzee, 2002), and many of them are vectors of parasites causing disease in humans and other vertebrates. Several of the vectors prefer to breed in very small containers such as old tyres, pots, and in footprints, or in phytotelmata: pools of water held by plants, for instance in the axils of leaves. *Aedes aegypti* (Linnaeus, 1762) is the vector of yellow fever. While the disease does not occur in southern Africa, *Ae. aegypti* does. It breeds in tiny containers, and the eggs can withstand desiccation for years (Coetzee, 2002). Farnesi et al. (2015), comparing the degree of resistance to desiccation of eggs of three species of mosquito, have shown that

1063 eggs of Ae. Aegypti can survive in a dry form for months, those of Anopheles aquasalis 1064 Curry, 1932 for a day, and of *Culex quinquefasciatus* Say, 1823 for a few hours, these times 1065 relating to the thickness of the egg shell. The members of 'floodwater Aedes', which can be 1066 vectors of Rift Valley Fever, such as Ae. (Neomelaniconion) mcintoshi Huang, 1985, Ae. 1067 (Neo.) circumluteolus (Theobald, 1908), Aedes (Neo.) luridus Mcintosh, 1971, Aedes (Neo.) 1068 lineatopennis (Ludlow, 1905), Aedes (Ochlerotatus) caballus (Theobald, 1912) and Aedes 1069 (Och.) juppi Mcintosh, 1973, aestivate as eggs for up to 20 months (Jupp et al., 1980). 1070 Several members of the genus *Anopheles* are vectors of malarial parasites. Although some 1071 anophelines prefer to oviposit in temporary waters, neither the eggs nor the larvae are 1072 resistant to desiccation. First-instar larvae of An. gambiae s.l. are said to be amphibious 1073 (Miller et al., 2007), in that they can survive in the film of water on the surface of mud. Both 1074 An. gambiae s.s. and An. arabiensis Patton, 1905 (two South African vectors of malaria) 1075 prefer sunny temporary pools and puddles, the major adaptation to this biotope being very 1076 rapid larval development. Mosquitoes appear to overwinter as larvae in permanent waters 1077 (Appleton et al., 1995). 1078 A few species of other dipteran families are known to occur in temporary waters in 1079 southern Africa, but very little is known about their systematics or their biology. We 1080 hypothesise that those occurring in the very arid western areas, such as the Namib Desert, 1081 have some specific adaptations that allow them to persist in areas where rain pools are the 1082 only surface freshwaters and these may fill only every few years. 1083 Insecta: Coleoptera 1084 Worldwide, beetles are one of the most abundant and speciose macroinvertebrate groups in 1085 inland waters, occurring across the entire hydrological spectrum, from the smallest puddles 1086 and phytotelmata, to large lakes and rivers. Water beetles are not a single taxonomic group, 1087 and instead result from around 15-20 transitions between land and water during the evolutionary history of the Coleoptera (Jäch & Balke, 2008; Mckenna et al., 2015; Short, 1088 1089 2018). Beetles are diverse and frequent inhabitants of temporary waterbodies, where 1090 carnivorous taxa such as diving beetles (Dytiscidae) are often the top predators. Temporary 1091 pool inhabitants are typically a mix of specialist and more generalist species, some of which 1092 may only occur there temporarily as adults. In many cases, specialising in temporary pools

may have arisen as a life-history strategy to evade fish predation, and in the northern

hemisphere some taxa characteristic of temporary ponds (e.g. *Agabus labiatus* (Brahm, 1790)) are also found in permanent, acidic, fish-free sites (Foster et al., 2016). Unlike most freshwater insect groups, the majority of water beetles are aquatic as both larvae and adults, but in many cases, the precise ecology of these life-history stages differs to some extent. Some beetles utilise temporary waters primarily for reproduction, with adults occupying different habitats. In the Mediterranean, *Acilius duvergeri* Gobert, 1874 for example, breeding occurs in temporary lagoons, where larvae feed on microcrustacea in open water (a strategy which would make them vulnerable to fish predation in permanent waters), whilst adults occupy permanent ponds during summer drought (Dettner, 1982).

Temporary pond water beetles have a variety of strategies to survive during the dry phase, reflecting the diverse origins of aquatic beetles themselves, and the repeated evolution of temporary water specialists from within these aquatic lineages. Temporary water species may have shorter larval duration compared to their permanent water relatives, allowing development to be completed during relatively short hydroperiods (e.g. Peters, 1972), and a range of approaches exists to surviving dry phases, including migration to permanent waters or adult/egg diapause *in situ* (e.g. Nilsson, 1986a, b, c). The most temporary of water bodies appear to be devoid of breeding water beetles, suggesting that the extent to which larval duration can be truncated may be limited. In addition, beetle larvae do not seem to have the same resistance to desiccation as many temporary-water Crustacea (see above), but do have the advantage that adults of most species can fly between habitats (e.g. Bilton et al., 2001).

In southern Africa, members of a number of beetle families can be found in temporary water bodies, although there are no formal publications dealing with their biology and ecology. As a consequence, the following information is based on Bilton (pers. obs.), unless otherwise stated. The faunas of vegetated temporary pools and rock pools appear to be quite distinct, and in addition brackish water bodies have their own specialised beetle fauna. Haliplidae are relatively rare in southern Africa, and members of *Haliplus* and *Peltodytes* are mostly found in larger, more permanent waters. The rare Cape endemic, *Algophilus lathridioides* Zimmermann, 1924 is poorly known ecologically, but appears to be a specialist of slightly mineralised waters. The only recent record of this beetle is from the Berg River floodplain close to Hopefield, where it occurs abundantly in slightly brackish temporary flood pools in grassland. *Algophilus* is wingless, and is likely to be dispersed during floods, having been found in floodwater along the Berg River in spring.

1126 Whereas the Noteridae are largely permanent water inhabitants, a number of Dytiscidae 1127 are temporary pond specialists, and others are found there occasionally or temporarily as 1128 adults. Canthyporus species occur across a spectrum of freshwater habitats, including 1129 seepages, streams and ponds, both permanent and temporary (Biström & Nilsson, 2006; 1130 Bilton et al., 2015). This genus is particularly diverse in the Cape, where most of its species 1131 are endemic. Many of the species in this genus, including C. aenigmaticus Biström & Nilsson 1132 2006 and C. lateralis (Boheman, 1848), are characteristic inhabitants of small mountain rock 1133 pools, where both adults and presumed larvae can be found. Some inhabited pools form parts 1134 of temporary stream/seepage systems during periods of high rainfall, whereas others 1135 constitute completely isolated basins. Cape vernal pools are also inhabited by Canthyporus, 1136 including the relatively generalist C. hottentottus (Gemminger & Harold, 1868) and C. 1137 petulans Guignot, 1951, as well as C. exilis (Boheman, 1848), a species otherwise found in 1138 temporary seeps and marshes. Besides the widespread genus *Hyphydrus*, the South African 1139 Hyphydrini includes five endemic genera (see Ribera & Balke, 2007), three of which occur in 1140 temporary ponds. Darwinhydrus solidus Sharp is found from the Cederberg to Port Elizabeth, 1141 in both permanent and temporary waters, but is most frequent in densely vegetated Cape 1142 vernal pools. *Primospes suturalis* Sharp, 1882 is largely restricted to the far southwest of the 1143 Western Cape, and occurs in temporary pools and rivers, and two *Hydropeplus* species are 1144 inhabitants of both stream pools and temporary lentic pools, both vegetated and rock pools. 1145 Species of the genus *Rhantus* can also be found in vegetated temporary ponds, as both adults 1146 and presumed larvae, as can the phylogenetically isolated Cape endemic Caperhantus 1147 cicurius (Fabricius, 1787) (Bilton, 2017). The Critically Endangered Cape endemic 1148 Capelatus prykei Turner & Bilton 2015 has been found in vernal pools on the Cape Flats 1149 (Bilton et al., 2015). 1150 To date, only a single species of the largely Holarctic genus *Helophorus* has been reported 1151 from southern Africa. Helophorus aethiops Balfour-Browne, 1954 is found throughout much 1152 of the winter rainfall zone, and is common in a variety of temporary pools, particularly 1153 shallow sites with exposed silt or sand substrates. The species-level taxonomy of southern 1154 African Hydrochus remains incompletely resolved, but a number of species are found in 1155 vegetated temporary ponds, as can many members of the Hydrophilidae, including *Enochrus* 1156 hartmanni Hebauer, 1998; Enochrus (Methydrus) spp.; Helochares spp; Paracymus spp; 1157 Berosus spp. and Limnoxenus sjoestedti Knisch, 1924, the latter being restricted to the South 1158 African Cape (Short & Liebherr, 2007).

The regional hydraenid beetle fauna is one of the most diverse on earth, most species in the region being Cape endemics. Here the family also attains its maximum ecological and morphological diversity, with both aquatic and (semi)terrestrial representatives (e.g. Perkins & Balfour-Browne, 1994). Species of the genus *Prosthetops*, which includes some of the largest hydraenids on earth, are characteristic inhabitants of temporary rock pools, where larvae and adults can be abundant, grazing biofilms. Bilton (2013) noted that large larvae of *P. wolfbergensis* Bilton, 2013 became active underwater in previously dry rock pools one day after rains, suggesting some desiccation resistance mechanism in these insects. During dry periods, *Prosthetops* adults can be found in rocky streams, although this does not appear to be their breeding habitat. Temporary vegetated freshwater ponds are inhabited by a number of species of *Hydraena* (*Hydraenopsis*), *Ochthebius* and *Parhydraena*, whilst temporary saline pools are home to species of *Ochthebius*, including *O. capicola* (Péringuey, 1892), a specialist of supralittoral coastal rock pools (Sabatelli et al., 2013), a habitat independently colonised by members of this genus on a number of occasions worldwide (Sabatelli et al., 2016).

Discussion

Much of southern Africa is characterised by a semi-arid or arid climate (Davis, 2011) and small temporary wetlands, though not widely acknowledged, are one of the most abundant and ubiquitous aquatic features of such landscapes (Silberbauer & King, 1991; Brendonck & Williams, 2000; Williams, 2006). The invertebrates found in these wetlands, whilst not widely acknowledged, constitute a significant proportion of the total aquatic biodiversity of the region. Whilst some recent research has highlighted the ecological and biodiversity importance of temporary wetland invertebrates (e.g. Spencer et al., 2002; Nicolet et al., 2004; Urban, 2004; Jeffries, 2005; Williams, 2006; Vanschoenwinkel et al., 2007; Ruhí et al., 2013; Batzer & Boix, 2016; Kneitel, 2016; Dalu et al., 2017a), baseline biodiversity data on these faunas are often lacking. Here we present such a synthesis for southern Africa, in the hope that this will aid further ecological investigations of temporary wetlands in the region.

A clear finding of this work is that, with the exception of a few groups (e.g. gastropod molluscs, odonates and certain dipteran families that act as disease vectors), we are often unable to reliably name many of the invertebrate organisms inhabiting these waterbodies. This becomes most obvious for the 'lower' invertebrate groups covered in this review, which is perhaps not surprising given their small size and the practical difficulties which may be

associated with collection and identification. For some of these groups, notably gastrotrichs, bryozoans and tardigrades, there is very little information about the freshwater species that occur throughout the region, including temporary wetlands. Although there is better information in this regard for the Northern Hemisphere, the 'lower' invertebrate phyla of freshwater environments have been relatively neglected worldwide (see Balian et al., 2008). Perhaps more surprisingly, the status quo reveals that in some cases we cannot put reliable names on species for groups represented by larger and more 'charismastic' organisms such as the 'clam shrimps' (Laevicaudata, Cyclestherida and Spinicaudata, but see Rogers et al., 2012 for revision of the family Limnadiidae), ephemeropterans and many of the hemipterans from temporary freshwater wetlands of the region. Harpacticoid copepods are another example of a taxon completely unstudied in the region, even from permanent freshwater systems.

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To get a better hold on patterns of richness and endemism, one first needs to know what species are present in these wetland systems. A case in point is the Cladocera, which have traditionally been considered to contain few endemics in the region (Frey, 1993; Korovchinsky, 2006; Smirnov, 2008). More recent work suggests that the apparent low endemism is most likely an artefact of the limited state of knowledge of the fauna, rather than genuinely low endemism (Van Damme et al., 2013). Thus, statements on patterns of richness and endemism are of a preliminary nature at present for many of the invertebrate groups. A few such patterns are worthy of comment here. First, only for a few of the groups is evidence presented that they reach higher richness and/or endemicity in temporary wetlands than for their permanent wetland counterparts. These taxa include large branchiopods, ostracods, copepods, cladocerans, certain diperans (notably culicids) and coleopterans (some dytiscids). Thus, for the vast majority of freshwater invertebrate taxa in the region, permanent waterbodies would appear to hold a greater richness and/or endemicity than temporary ones. Despite this, temporary wetlands support unique and specialised taxa. For instance, the temporary wetland ostracod fauna of northern and central Namibia and Botswana has a generic endemicity rivalled worldwide only by East African paleolakes (Martens, 2001). Another feature of the ostracod fauna is the specific diversity in the family Lymnocytheridae for rock pools of the Drakensberg (SA), which is unmatched anywhere else in the world (Martens et al., 1998). Southern Africa is considered one of the world's hotspots of large branchiopod diversity (Tuytens et al., 2015), almost all of which is housed in temporary wetlands. When considering the region's area, the anostracan fauna is one of the richest worldwide and also has a very high endemicity (~80% at the time of the review of Hamer &

Brendonck, 1997). Other than for the anostracans and ostracods, further commentary on which of the major invertebrate groups covered in this review stand out globally for their richness/endemism is limited due to the taxonomic uncertainty associated with most of the groups in the region, as highlighted above.

Identifications to genus and species level of specimens mentioned in this review are mostly based on morphological characteristics (unless specifically mentioned otherwise) and accuracy greatly depends on reliable identification keys and taxonomic expertise, which is considered a scarce skill in southern Africa (Hamer, 2013). The few region-specific keys that exist are in need of updating (Barber-James & Gattolliat, 2012). Species identification using molecular systematics has only become prominent in the region recently (da Silva & Willows-Munro, 2016) and will greatly contribute to more accurate identification of species recorded in temporary wetlands in future.

Another general trend picked up from this review is that virtually all of the major invertebrate groups possess some sort of adaptation for survival in temporary wetlands. Although only certain taxa can truly withstand desiccation (e.g. branchiopods, ostracods and copepods), many taxa have dispersal mechanisms which allow them to leave the waterbody before it dries up (e.g. leeches, water mites and odonates), or are able to survive the drought phase as adults (e.g. dipterans). Many of the taxa inhabiting these environments are habitat generalists that may opportunistically exploit temporary waterbodies for the advantages afforded by these environments, such as reduced levels of predation and competition (although this notion is in itself controversial, see Brendonck et al., 2002). A case in point is the Odonata and Hemiptera, which are common and widespread in temporary wetlands of the region, but tend to be opportunistic colonisers of these environments, depending on dispersal for survival and having large home ranges, and thus are generally not threatened. This generalistic nature of many of the inhabitants of temporary wetlands in the region is hypothesized as one of the reasons why many of the taxa do not display high levels of endemism. There are certain exceptions, such as for rock pools of the Drakensberg in the east of the region, and the lowland pools of the Western Cape province, both of which are hotspots of ostracod and cladoceran endemism and richness (Martens et al., 1998; Martens, 2001; de Moor & Day, 2013; Van Damme et al., 2013).

In terms of the best known of the temporary wetland specialists, 'fairy shrimp' (anostracan) species richness is depicted as an array of hotspots throughout the region, not

being centered on any particular area and does not appear to be governed by any particular climatic factor (Hamer & Brendonck, 1997). The large branchiopod crustaceans show the greatest potential as 'flagships' for conservation of temporary wetlands in the region, given their relatively large size and uniqueness in terms of adaptations to life in the temporary aquatic realm. The odonates, molluscs and anostracans are the only groups covered in this review to have an appreciable amount of IUCN Red List information in the region. The freshwater molluscs appear to be have few threatened species in the region (10 species, 8.6% of the fauna, Kristensen et al., 2009). Anostracan Red List information is still far from comprehensive. For instance, De Roeck et al. (2007) reported that only two of the 14 anostracan species known to occur in the Western Cape province of SA were listed by IUCN and that insufficient data were available to determine the IUCN Red Data Category of six of these species. Of the 46 anostracan species reported for southern Africa by Hamer & Brendonck (1997), two were considered by IUCN (1994) as 'Critically Endangered', four were 'Endangered', three were 'Vulnerable' and 24 were 'Least Concern', whilst data were deficient for the categorisation of 13 species. As is the trend worldwide, the anostracans of the region are threatened by agriculture and urbanization, due to both indirect (pollution and pesticides) and direct (physical) habitat alteration arising from these activities (Brendonck et al., 2008). These factors are thought to have driven the extinction of Streptocephalus gracilis Sars, 1898 and *Branchipodopsis karroensis* Barnard, 1929 (De Roeck et al., 2007).

We hypothesize that temporary wetland specialists such as the branchiopods (including cladocerans), ostracods and copepods, which tend to be more habitat-restricted and less capable of escaping degradation, are more likely to be affected by human impacts than for taxa that are able to easily disperse to other more favourable areas (e.g. dipterans, hemipterans, odonates and coleopterans). The sensitivity of the region's temporary wetland invertebrates to human impacts such as pollution and habitat alteration is a topic still under investigation, with some evidence indicating a resilient fauna (Bird et al., 2013), whilst other evidence is in favour of a more sensitive fauna (Bird & Day, 2016), especially when effects are combined with climate change impacts (Dalu et al., 2017a). Whatever the case may be, what is certain is that temporary wetland habitats worlwide have been degraded and destroyed at an alarming rate (Semlitsch & Bodie, 1998), and various studies indicate that the situation is no different in southern Africa (De Roeck et al., 2007; Bird & Day, 2014; Dalu et al., 2017a).

In terms of the way forward, we cannot expect to properly understand and conserve temporary wetlands if we don't know what lives in them. As things stand, this is certainly the case in southern Africa in that we lack updated taxonomic information for many groups that inhabit these systems. This review shows that we have a base of knowledge on which to work, but the now famous stagnation of taxonomic expertise both locally and abroad (see Godfray, 2002; Wheeler et al., 2004; Agnarsson & Kuntner, 2007; Bebber et al., 2014) is a major threat hindering the further progression of this knowledge. In addition to providing a synthesis for southern Africa, this work highlights the dearth of fundamental taxonomic information on the invertebrate fauna of temporary wetlands in the region, and constitutes a plea for more foundational taxonomic work to be initiated.

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